Biocatalytic oxidative cleavage of alkenes using novel metal-dependent aromatic dioxygenases

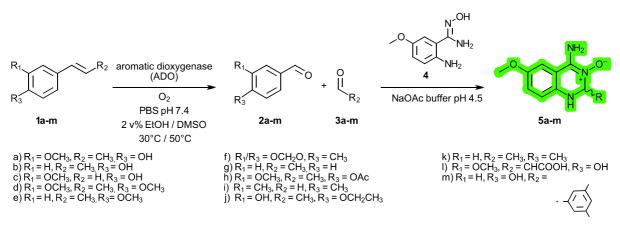
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Scheme 1: Left: Oxidative cleavage of **1** by aromatic dioxygenases (ADOs) to the corresponding aldehydes (**2**, **3**). Right: Detection of the formed aldehydes via ABAO-assay using 2-aminobenzamidoxime derivate **4**. The resulting quinazolines **5** exhibit UV absorption and fluorescence properties.

Oxidative cleavage of alkenes can be used to obtain carbonyl compounds, valuable building blocks in various areas, including food, flavor, and the pharmaceutical industry.¹ While this can be done *via* chemical approaches, like ozonolysis², we are interested in milder and safer enzymatic approaches using novel metal-dependent aromatic dioxygenases (ADOs). In the presence of oxygen, these enzymes enable the oxidative cleavage of substrates such as isoeugenol (1a) to the corresponding aldehydes (Scheme 1). For the substrate screening, HPLC, GC, or a pooling approach based on the reported ABAO-assay³ are used. The latter is a photometric/ fluorometric assay for the detection of the aldehydes. It is based on the rapid reaction of aldehydes with 2aminobenzamidoxime (ABAO), forming quinazolines 5 that exhibit UV absorption and fluorescence properties. Using this assay, biotransformations with several pooled substrates are analyzed by detecting the photometric/fluorometric responses. So far, substrates such as isoeugenol (1a), hydroxyanethole (1b), 4-vinylguaiacol (1c), and resveratrol (1m) were found to be successfully converted to the respective aldehydes in a whole-cell system reaction. Among the tested enzymes, especially the MapADO from Moesziomyces aphidis, showed promising properties and converted 10 mM isoeugenol to vanillin in a whole cell approach within 1 h. Furthermore, the crystal structure of MapADO could be obtained, and several rational mutants were generated. So far, the screening revealed that the para-hydroxy group is essential for substrate acceptance.

⁽¹⁾ Kazimírová, V.; Rebroš, M. Production of Aldehydes by Biocatalysis. *Int. J. Mol. Sci.* **2021**, 22 (9), 4949.

⁽²⁾ Fisher, T. J.; Dussault, P. H. Alkene ozonolysis. *Tetrahedron* 2017, 73 (30), 4233.

⁽³⁾ Ressmann, A. K.; Schwendenwein, D.; Leonhartsberger, S.; Mihovilovic, M. D.; Bornscheuer, U. T.; Winkler, M.; Rudroff, F. Substrate-Independent High-Throughput Assay for the Quantification of Aldehydes. *Adv. Synth. Catal.* **2019**, *361* (11), 2538.